



## Research

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# Oxygen, temperature and the deep-marine stenothermal cradle of Ediacaran evolution

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Ediacaran fossils document the early evolution of complex megascopic life, contemporaneous with geochemical evidence for widespread marine anoxia. These data suggest early animals experienced frequent hypoxia. Research has thus focused on the concentration of molecular oxygen ( $O_2$ ) required by early animals, while also considering the impacts of climate. One model, the Cold Cradle hypothesis, proposed the Ediacaran biota originated in cold, shallow-water environments owing to increased  $O_2$  solubility. First, we demonstrate using principles of gas exchange that temperature does have a critical role in governing the bioavailability of  $O_2$ —but in cooler water the supply of  $O_2$  is actually lower. Second, the fossil record suggests the Ediacara biota initially occur approximately 571 Ma in deep-water facies, before appearing in shelf environments approximately 555 Ma. We propose an ecophysiological underpinning for this pattern. By combining oceanographic data with new respirometry experiments we show that in the shallow mixed layer where seasonal temperatures fluctuate widely, thermal and partial pressure ( $pO_2$ ) effects are highly synergistic. The result is that temperature change away from species-specific optima impairs tolerance to low  $pO_2$ . We hypothesize that deep and particularly stenothermal (narrow temperature range) environments in the Ediacaran ocean were a physiological refuge from the synergistic effects of temperature and low  $pO_2$ .

## 1. Introduction

The role of marine oxygenation as it pertains to early animal evolution is a fundamental question in deep Earth history. Interdisciplinary research spanning palaeontology, geochemistry, and molecular biology increasingly tie changes in Earth's surface environment to the emergence and subsequent radiation of animals across the Neoproterozoic-early Palaeozoic transition approximately 800–500 million years ago (Ma). This interval is marked by evidence for extreme climate fluctuations and biogeochemical perturbations, including two long-lasting glaciation events (i.e. snowball earth glaciations) during the Cryogenian Period *ca* 720–635 Ma [1], and extremely positive carbonate carbon isotope records punctuated by negative excursions [2]. The first large, morphologically complex fossils do not appear in the fossil record until the Ediacaran Period *ca* 635–541 Ma [3]. Multi-proxy geochemical evidence suggests early animal evolution occurred against a backdrop of widespread marine subsurface anoxia [4–6]. While the absolute amount of oxygen ( $O_2$ ) as a percentage of present atmospheric levels (PAL) through the Neoproterozoic are contentious, researchers have traditionally considered levels of 1–10% PAL to be most likely [7,8]. During the Ediacaran and early Cambrian, redox sensitive trace metal enrichments suggest transient oxygenation events during that time [9,10]. However, regional and stratigraphic inconsistencies in the pattern of these enrichments and several other lines of evidence indicate any oxygenation must have been relatively muted or short-lived [5,6,11–14]. There is therefore emerging consensus that early animals encountered persistent and severely

low  $O_2$  partial pressures ( $pO_2$ ), which would have had profound effects on the spatial distribution of metabolically-viable habitats [15].

For decades, these low levels of marine  $O_2$  have been opined [16–19], albeit controversially [20], as the environmental barrier to early animal evolution. However, given the striking climatic fluctuations of the Neoproterozoic,  $O_2$  is not the only environmental influence that has been considered. The stratigraphic occurrence of highly diverse, shallow-water shelfal ‘White Sea’ fossil assemblages *ca* 560–550 Ma [3] in exclusively siliciclastic sediments has been viewed as an indication of habitation in cold-water environments (at latitudes above low-latitude carbonate belts) [21]. To the extent that the Ediacara biota represent animals, this ‘Cold Cradle’ model of evolution posited that the greater gas solubility of  $O_2$ , as well as the sluggish remineralization of nutrients by prokaryotes in cold, shallow, high-latitude waters allowed metazoan ecosystems to flourish in the later Ediacaran.

Detailed palaeontological and geochronological studies now indicate that the oldest non-algal megascopic fossil assemblage is not the White Sea assemblage, but rather is the so-called ‘Avalon assemblage’ [22–25]. This community is primarily dominated by morphologically complex soft-bodied benthic frondose fossils belonging to two recognized groups, the *Arborea* and *Rangia*. While there are phylogenetic issues with assigning Ediacaran fronds to the Metazoa ([26], but see [27]), Avalonian assemblages also contain other fossils more likely to be animals, including sponges [28] and body- and trace-fossil evidence for eumetazoan cnidarians [23]. These fossils appear in the middle Ediacaran *ca* 571 Ma in deep-water, aphotic slope and basinal facies [22–24,29], where they are found *in situ*, buried by ash beds or rapidly deposited sediments. The inferred deep-water depositional environment is supported by expansive, kilometre-scale stratigraphic sections of uninterrupted turbidites displaying thick  $T_{C-E}$  and  $T_{D-E}$  Bouma subdivisions, contour-parallel bottom currents, and no evidence for wave-generated sedimentary structures [30]. These stratigraphically oldest deep-water, morphologically complex fossils are found in multiple sedimentary basins worldwide including England, Newfoundland, and the Mackenzie and Wernecke Mountains of northwestern Canada. By contrast, Ediacara biota are absent from shallow-water environments on the shelf until approximately 560–555 Ma, often after the globally recognized Shuram carbon isotope excursion [31–33].

The Ediacaran fossil record therefore displays a puzzling pattern. For approximately 15 Myr large, morphologically complex eukaryotes and animals only inhabited deep-water settings. This observation is at odds with palaeontological meta-analyses which show that onshore to offshore macro-evolutionary patterns predominate across multiple intervals in the later Phanerozoic (541 Ma-present) fossil record [34]. Might deep-water settings have provided a kind of physiological refugia for early metazoans in a generally low  $pO_2$  global ocean? Given that both deep- and high-latitude water masses are colder than shallow-water counterparts at low-latitudes, what ecophysiological or oceanographic differences exist between the two environments, and how do they fit within the context of the Cold Cradle hypothesis? Lastly, as Ediacaran communities did eventually radiate onto the shelf to inhabit shallow-water environments, what stressors might these ecosystems have faced? To shed light on these questions, we apply an oxygen supply index (OSI, table 1)

to determine how temperature can govern  $O_2$  supply to animals at oceanographic scales. We then present new experimental respirometry data that illustrate how temperature dynamically affects the absolute  $pO_2$  tolerance of marine ectotherms. Lastly, we integrate these two approaches to re-examine bathymetric patterns within the Ediacaran fossil record in an ecophysiological context.

## 2. Background and previous work

### (a) Oxygen bioavailability in aquatic settings

The challenges of aquatic respiratory gas exchange are well known. In water at standard temperature and pressure there is approximately 30 times less  $O_2$  than in the atmosphere by concentration, and  $O_2$  diffuses approximately  $2.4 \times 10^5$  times slower through water than air [35]. Furthermore, both temperature and salinity independently impact the solubility of  $O_2$  in water owing to their effects on the Henry gas coefficient. This results in seawater containing on average 25% less  $O_2$  than freshwater at a given temperature [36]. Owing to these physical constraints, limitations in environmental  $O_2$  supply have profound physiological impacts on aquatic animals in the modern ocean.

Respiratory  $O_2$  exchange cannot simply be discussed interchangeably with units of  $pO_2$  or solubility. Instead, the product of  $pO_2$ , solubility, and diffusivity potential together represent the flux of  $O_2$  that can be transferred from the environment into a respiring organism. This relationship inherently describes Fick’s first law of diffusion, which expresses partial pressure as proportional to its concentration gradient [37]. Putting aside organism-specific differences in surface area to volume ratios, respiratory structures (e.g. gills, pigments), or differences in pumping [38], aquatic animals can only extract  $O_2$  from the water column at an absolute rate proportional to environmental availability. This relationship has been expressed in freshwater environments as the OSI [39] (expressed here in  $\mu\text{mol kg}^{-1} \text{matm m}^2 \text{s}^{-1} \times 10^{-5}$ ):

$$OSI \propto \alpha_{O_2} \cdot DO_2 \cdot pO_2, \quad (2.1)$$

where  $\alpha_{O_2}$  is the solubility of  $O_2$  in water ( $\text{mol m}^{-3} \text{Pa}^{-1}$ ),  $DO_2$  is the diffusivity of  $O_2$  in water ( $\text{m}^2 \text{s}^{-1} \times 10^{-9}$ ), and  $pO_2$  is the partial pressure of  $O_2$  in water (matm).

### (b) Respiration physiology and temperature

The ability to avoid hypoxia in low  $O_2$  conditions represents a physiological balance between  $O_2$  supply and demand. Temperature affects both sides of this equation [40], and for aquatic ectothermic animals, the inability to control body temperature results in metabolic rates which change significantly with ambient temperature. All ectotherms exhibit a  $Q_{10}$  rate coefficient in which a  $10^\circ\text{C}$  increase in body temperature raises metabolic rate by a factor of approximately 2–3, or about 8% per degree Celsius [41]. Despite numerous processes at the whole-organism, tissue, and enzymatic levels to partially offset this effect, these strategies are invariably not fully effective at countering the effects of temperature on metabolic rate [42]. Marine ectotherms consequently display a thermal performance curve (table 1) which often represents an organism’s relative fitness across its natural environmental temperature range [43]. The thermal performance curve reflects the effects of low temperature on  $O_2$  supply and ventilation costs, and high temperatures on

**Table 1.** Summary definitions of key terms.

term	abbreviation	definition
oxygen supply index	OSI	a term which measures the rate at which oxygen can transfer from the water column into an animal by integrating the partial pressure and diffusivity of oxygen within the water as well as its solubility
thermal performance curve	TPC	the thermal performance curve represents an animal's fitness due in part to thermal tolerance characteristics and temperature dependant effects on physiological and biochemical functions (e.g. fecundacy, growth, metabolic rate). Often it reflects the natural environmental temperature range of the species, that is its thermal window
thermal optimum	$T_{opt}$	maximum performance occurs here at the peak of the TPC, often at intermediate temperature and represents maximum aerobic scope, that is the greatest difference between MMR and SMR
standard metabolic rate	SMR	the minimum metabolic rate that supports basic maintenance requiremens of an organism while at rest and fasting. In ectotherms this rate is temperature sensitive
maximum metabolic rate	MMR	the maximum metabolic rate achieved by an organism during unsustainable physical activity that is limited by aerobic capacity
oxygen- and capacity-limited thermal tolerance	OCLTT	a concept which describes thermally-induced hypoxemia (low levels of oxygen in blood or tissues) at both ends of an animal's thermal window due to thermal effects imparted on oxygen bioavailability, metabolic demand, and ventilatory capacity to supply enough oxygen to meet these metabolic requirements
critical $O_2$ level	$[O_2]^{crit}$	this is the critical oxygen level below which standard metabolic rate can no longer be maintained aerobically. At this point, oxygen demand is greater than the animal's capacity to supply oxygen and anaerobic metabolic pathways begin operating

enzyme instability and metabolic demand. In respiratory physiology terms, the thermal performance curve inherently represents aerobic scope, because ATP yield from aerobic respiration is dramatically higher (approx. 15×) than any anaerobic glycolytic pathway. Scope is the instantaneous proportion of metabolic power available to an organism after its basal maintenance costs are met, and can be used to invest in growth, reproduction, predation and defence, and other fitness-related functions [44].

The co-limiting effects of temperature and low  $pO_2$  on aerobic scope can result in the metabolic demands of an animal exceeding its capacity for  $O_2$  supply [45]. This phenomenon is referred to as oxygen- and capacity-limited thermal tolerance (OCLTT, table 1) [46]. At its most basic level, OCLTT represents a reduction of relative aerobic scope at temperatures both above and below  $T_{opt}$  (table 1). Above  $T_{opt}$  standard metabolic rate (the lowest rate of  $O_2$  consumption required for maintenance, table 1) makes up a greater proportion of aerobic scope owing to  $Q_{10}$  effects (figure 1) [45]. At high enough temperatures, maximum metabolic rate (table 1) can also begin to decrease owing to heat-induced damage to biochemical systems and organ (e.g. heart) failure [47,48]. Combined, warming above  $T_{opt}$  causes a decrease in aerobic performance and lowers  $pO_2$  tolerance, causing survivorship to greatly decline [49]. At temperatures below  $T_{opt}$  maximum metabolic rate becomes increasingly inhibited owing to decreasing  $O_2$  bioavailability, and reduced aerobic capacity owing to kinetic reduction in circulatory and ventilatory performance. This results in a net reduction of aerobic scope despite lower metabolic rates, thus limiting low  $pO_2$  tolerance at the cold end of the spectrum as well (figure 1)

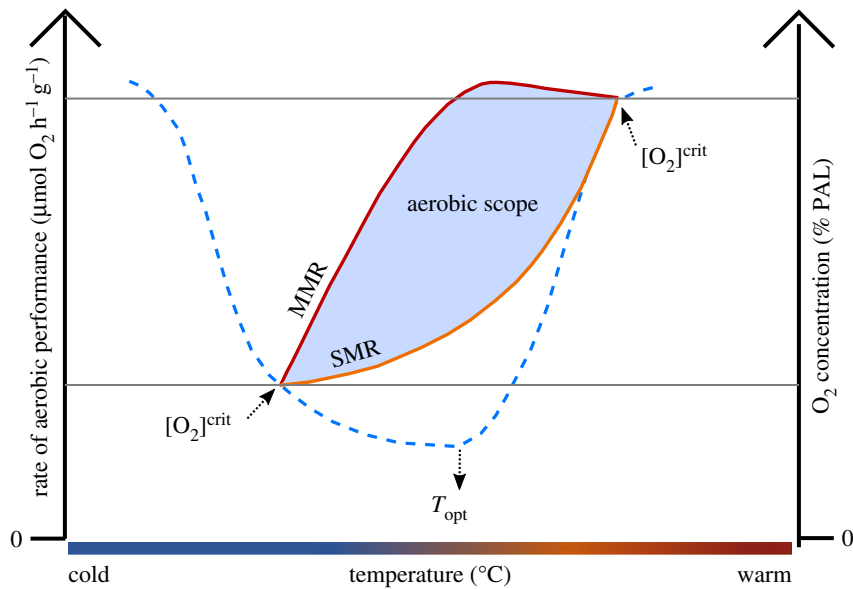
[47,48,50]. Lastly, at both ends of the thermal performance curve, low ambient  $pO_2$  not only drops maximum metabolic rate (causing a reduction in the range of thermal tolerance by way of reducing overall scope), but also increases the proportion of standard metabolic rate used for ventilation, as animals must exchange more water over their respiratory surfaces in order to extract the same amount of  $O_2$  [51,52].

The OCLTT relationship can be determined for a given animal by measuring what is known as the critical  $O_2$  concentration ( $[O_2]^{crit}$ , table 1), or alternatively expressed in partial pressure ( $pO_2^{crit}$ ), across its natural temperature range.  $[O_2]^{crit}$  represents the  $O_2$  level below which an organism is no longer able to maintain its standard metabolic rate. At the  $[O_2]^{crit}$ , aerobic scope is therefore zero, and any further decrease in  $O_2$  saturation, or change in temperature further away from  $T_{opt}$  will result in ATP demand exceeding  $O_2$  supply. At a metabolic level, this deficit in aerobically generated ATP triggers the onset of fermentation. This shift in metabolic pathway dramatically changes respiration rate, allowing  $[O_2]^{crit}$  to be calculated using breakpoint analysis on the  $O_2$  draw-down curve generated during closed-system experimental respirometry (electronic supplementary material, figure S9).

### 3. Results

#### (a) Oceanographic controls on oxygen supply

OSI has been calculated in freshwater environments, but not in the global ocean. To illustrate the behaviour of OSI in the global ocean for the first time to our knowledge, dissolved  $O_2$ , salinity, density, temperature, and pressure data from



**Figure 1.** Conceptual model of how temperature and ambient  $O_2$  interact to constrain aerobic performance in ectothermic animals. Aerobic scope is defined as the difference of rates of aerobic performance (left axis), specifically between maximum metabolic rate, MMR, and standard metabolic rate, SMR. Tolerance to low  $O_2$  levels decreases at temperatures both below and above  $T_{opt}$ , as  $O_2$  supply capacity falls relative to  $O_2$  demand, resulting in  $[O_2]^{crit}$  occurring at progressively higher ambient  $O_2$  tensions (blue dashed line, right axis). Because  $T_{opt}$  represents maximum rate of aerobic performance (the greatest distance between MMR and SMR on the left axis), it also corresponds to the  $[O_2]^{crit}$  minimum (on the right axis). As low ambient  $O_2$  reduces aerobic scope by lowering MMR, it consequently narrows the thermal window of environmental  $O_2$  tolerance because any temperature-related increase in SMR or reduction in MMR will consequently take up a larger proportion of aerobic scope. Figure adapted from [46].

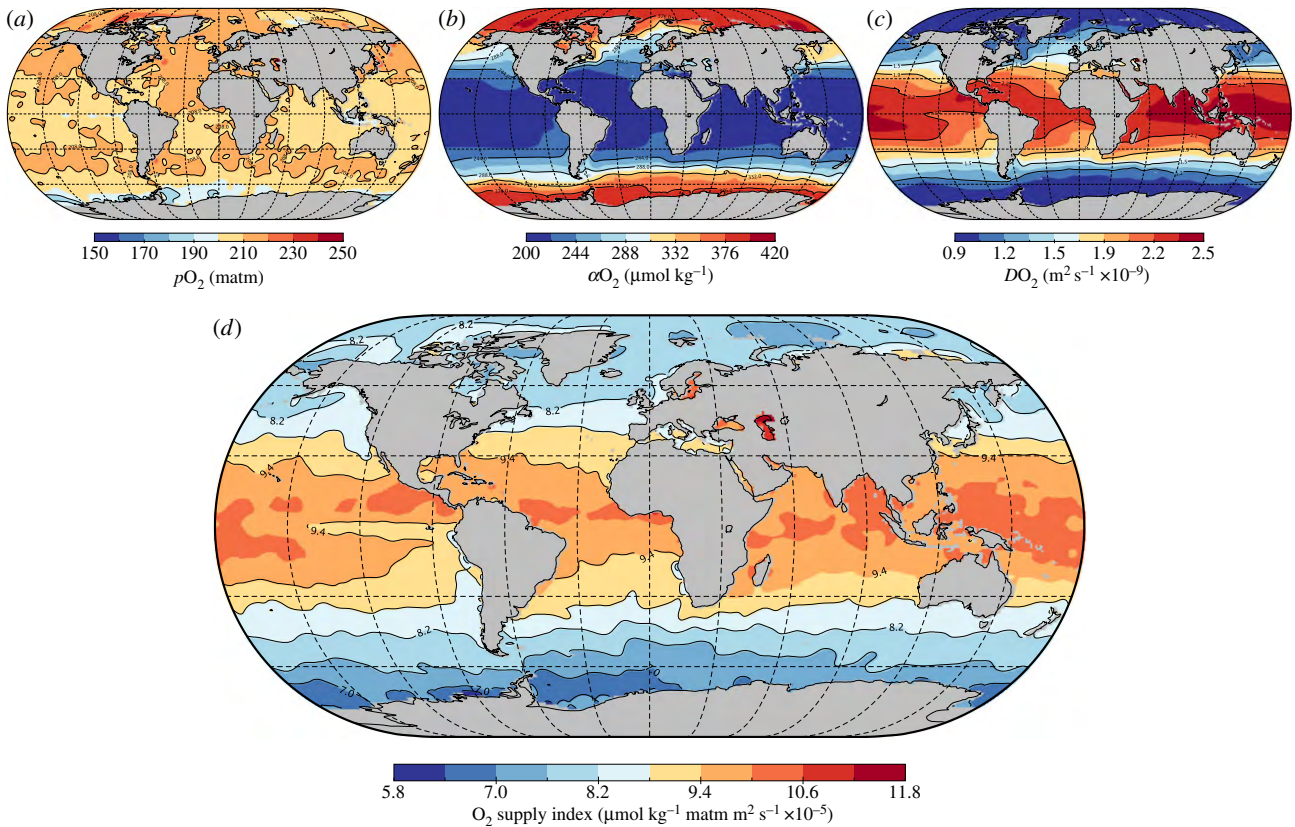
the National Oceanic and Atmospheric Administration World Ocean Atlas (NOAA WOA13 V2) were used to calculate *in situ*  $pO_2$ , diffusivity and solubility (electronic supplementary material). Temperature and salinity independently affect solubility. It is therefore possible for solubility to change dissolved  $O_2$  concentrations in the ocean without any adjustment in  $pO_2$ , such as across salinity or temperature gradients. Ultimately, the total amount of  $O_2$  that is present at a given location is dependent on not just solubility, but also on atmospheric  $pO_2$  and the saturation state of the water column (electronic supplementary material, figures S1–S3). At the sea surface, there is little variation in the  $pO_2$  of seawater with latitude owing to the limited change in saturation water vapour pressure across the normal range of marine water temperatures combined with the near constant atmospheric  $pO_2$  at sea level (figure 2; electronic supplementary material, figure S7; [53]). At depth though, increases in  $pO_2$  of approximately 14% per 1000 m depth occur owing to large increases in hydrostatic pressure (electronic supplementary material, figures S4–S6 and S8; [54]). However, because pressure increases the fugacity of  $O_2$  exponentially, it also has a reciprocal effect on the solubility coefficient in water owing to increased outgassing tendency (electronic supplementary material, figure S2a). This leads to little change in OSI past the thermocline unless the water mass is undersaturated with respect to  $O_2$  (electronic supplementary material, figures S1d and S4d). For instance, at depths between 200–1000 m in the modern ocean, undersaturation (lower than equilibrium dissolved  $O_2$  concentrations) driven by remineralization of organic matter plays a significant role in developing oxygen minimum zones (OMZs) on upwelling margins (electronic supplementary material, figure S6d; [55]).

Finally, the third component of OSI, diffusivity, is largely controlled by Brownian motion, as molecular  $O_2$  exists as a dissolved gas within seawater. As such, the capacity of  $O_2$  to diffuse through the liquid medium is heavily dependent

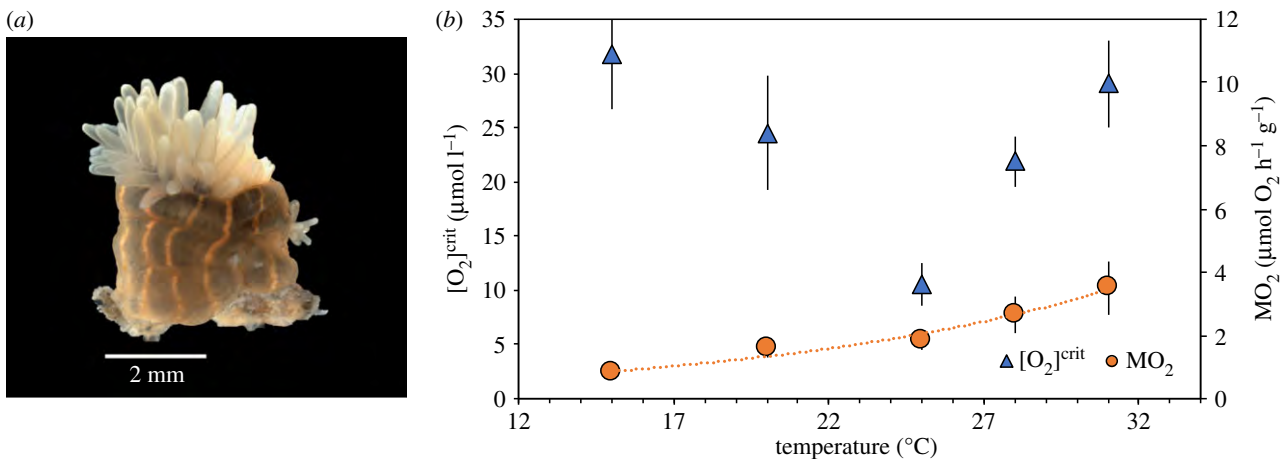
on the ratio of temperature to density of seawater, which together govern its kinematic viscosity (also known as momentum diffusivity) [56]. Given the wide thermal ranges and saline nature of seawater, it is critical to take diffusivity into account in studies of respiration physiology. However, this is not commonly done. Within the ocean, increases in viscosity owing to colder water temperatures at higher latitudes or greater depths overcome increases in solubility on OSI (electronic supplementary material, figures S4–S7). The key point here regarding  $O_2$  supply for respiration as determined from OSI is that cold marine waters at depth or high-latitudes in equilibrium saturation conditions have only 60–70% of the  $O_2$  supply available in shallow low-latitude regions (figure 2; electronic supplementary material, figures S4d and S7d). The physiological implications of this novel result are immediately apparent and probably far reaching, as cold waters in the global ocean are therefore much more difficult to respire in, despite having greater  $O_2$  solubility [39,57].

### (b) Aerobic respiration and temperature

To illustrate how  $[O_2]^{crit}$  varies with temperature, we conducted 86  $[O_2]^{crit}$  measurements on the intertidal anthozoan cnidarian *Diadumene lineata* (figure 3). This species of sea anemone was selected as intertidal organisms regularly encounter significant diurnal and seasonal temperature and  $pO_2$  fluctuations [58]. Furthermore, anemones possess a diploblastic body plan and therefore rely entirely on cutaneous diffusion of  $O_2$  into two epithelial tissue layers—the external ectoderm, and internal endoderm—for aerobic respiration. These layers are supported by a hydrostatic skeleton constructed of a gelatinous, metabolically-inert tissue called mesoglea. This makes the anemone body plan at least physiologically analogous, if not necessarily homologous, to many



**Figure 2.** Factors governing oxygen supply to animals. (a) Average annual partial pressure of O<sub>2</sub> ( $pO_2$ ) in the global ocean at surface. (b) Average annual solubility of O<sub>2</sub> ( $\alpha O_2$ ) in the global ocean at surface. Values increase with latitude owing to the thermal effects on Henry's solubility coefficient. (c) Average annual diffusivity of O<sub>2</sub> ( $DO_2$ ) in the global ocean at surface. (d) Average annual bioavailability of O<sub>2</sub> in the global ocean at surface, expressed using the oxygen supply index (OSI) [39]. Despite the increased solubility of O<sub>2</sub> in cold water, the kinematic viscosity also increases substantially, reducing the diffusivity of O<sub>2</sub> at a rate greater than the offsetting effect on solubility. As a result, the supply of O<sub>2</sub> to respiratory surfaces actually decreases approximately linearly as water becomes colder.



**Figure 3.** Variation of  $pO_2$  tolerance with temperature. (a) Intertidal anthozoan cnidarian *Diadumene lineata* (YPM IZ 077401) from the New England region of the western Atlantic. Image courtesy of E.A. Lazo-Wasem. (b)  $[O_2]^{crit}$  and  $MO_2$  data for 86 individuals of *D. lineata* binned into five experimental temperatures ( $\pm 0.3^\circ C$ ). Mean ( $\pm$ s.d.) standard O<sub>2</sub> consumption rate ( $MO_2$ ) increases consistently with temperature owing to the Arrhenius relationship ( $Q_{10} = 2.50$ ,  $n = 70$ ,  $R^2 = 0.96$ ). Absolute environmental O<sub>2</sub> tolerance ( $[O_2]^{crit}$ ) displays a bidirectional relationship ( $n = 86$ ,  $R^2 = 0.75$ ). Triangles represent average mean values and whiskers represent standard error. These data provide, to our knowledge, the first experimental support for physiological principles hypothesized to be universal for marine ectotherms [45,46], indicating these principles are applicable to questions regarding the deep-water origin of Ediacaran organisms. (Online version in colour.)

Avalonian Ediacaran organisms without circulatory or respiratory systems. The comparison may be even closer than analogue, as many workers interpret some morphologically complex Ediacaran fossils to in fact be total-group actinarian cnidarians [59–61].

Results of respirometry experiments demonstrate this taxon displays mass-normalized standard metabolic rates ( $MO_2$ ) that

increase predictably with temperature ( $Q_{10} = 2.50$ , figure 3). For  $[O_2]^{crit}$ , mean values take the form of a concave-up parabola, demonstrating the predicted bidirectional effects that temperature has on environmental O<sub>2</sub> tolerance. At a  $T_{opt}$  of approximately  $24^\circ C$ , aerobic scope is maximal and *D. lineata* is able to respire aerobically well into low  $pO_2$  levels ( $[O_2]^{crit}$  of  $10.4 \mu mol l^{-1}$  or 4.7% PAL). However, upon warming to

28°C,  $p\text{O}_2$  tolerance decreases significantly ( $[\text{O}_2]^{\text{crit}}$  of 21.9  $\mu\text{mol l}^{-1}$  or 10.4% PAL). Cooling to 20°C produces a similar decrease in tolerance ( $[\text{O}_2]^{\text{crit}}$  of 24.5  $\mu\text{mol l}^{-1}$  or 10.2% PAL). Such bi-directionality has been studied at the molecular level and predicted to occur at the organismal level [46–48,50,62], yet despite its purported generality as a physiological principle that affects all aquatic ectotherms [46], this relationship has not previously been demonstrated experimentally using respirometry. While the exact shape of the concave-up parabola and position of the thermal optimum almost certainly differs between organismal lineages in different environments and over evolutionary timescales, the bi-directional effects seen in these *D. lineata* respiratory data are likely to be universal. These novel experimental results demonstrate that hypoxia tolerance is not a single threshold value in dynamic, shallow-water marine environments, but rather is determined jointly from the effects of temperature on OSI and on an animal's supply capacity over metabolic demand.

## 4. Discussion

### (a) Seasonality and thermal tolerance in low $p\text{O}_2$ oceans

These physiological principles have significant use for understanding how low ambient  $\text{O}_2$  levels and climate might have synergistically affected early animal life. While focus on the absolute lower  $\text{O}_2$  limits for metazoan aerobic respiration [63,64] or critical thresholds on carnivory [65] provide important constraints on early animal ecosystems, the exogenous thermal environment in combination with  $p\text{O}_2$  probably governed the metabolic viability of habitats for early animal ecosystems. Critically, because ambient  $p\text{O}_2$  governs the size of aerobic scope, low  $p\text{O}_2$  narrows the thermal range over which aerobic respiration can occur. Mathematically, this can be thought of in terms of the area above a polynomial regression run through a plot of  $[\text{O}_2]^{\text{crit}}$  with respect to temperature for a given species, expressed as:

$$A = \left(\frac{2}{3}\right) \cdot b \cdot h, \quad (4.1)$$

where  $A$  is the area enclosed between the parabola and a chord intersecting the  $y$ -axis at an environmental  $\text{O}_2$  level. The length of this chord between the intercepts with the parabola is  $b$ , which represents the thermal range of aerobic respiration at a given  $\text{O}_2$  concentration.  $h$  is the height from the parabola vertex ( $[\text{O}_2]^{\text{crit}}$  at  $T_{\text{opt}}$ ) to the chord, and represents aerobic scope (figure 4a). Because  $A$  in equation (4.1) is related to the product of both  $\text{O}_2$  and temperature, any decrease in aerobic scope ( $h$ ) caused by lower ambient  $\text{O}_2$  concentrations will also narrow the thermal range of aerobic respiration ( $b$ ). For *D. lineata*, ambient  $p\text{O}_2$  at 14.5% PAL corresponds to a 16°C allowable range for aerobic respiration which meets minimum requirements for maintenance, but ambient  $p\text{O}_2$  at 10% PAL results in only a 9°C allowable range (figure 4). Thermal range continues decreasing to zero at a  $p\text{O}_2$  of approximately 5% PAL, where the animal can only respire aerobically at its  $T_{\text{opt}}$  of approximately 24°C. This relationship has significant implications when considering the impact of thermal variability in the Ediacaran ocean.  $p\text{O}_2$  levels of 10% PAL are on the high end of estimates for this interval, although in reality very little is concretely

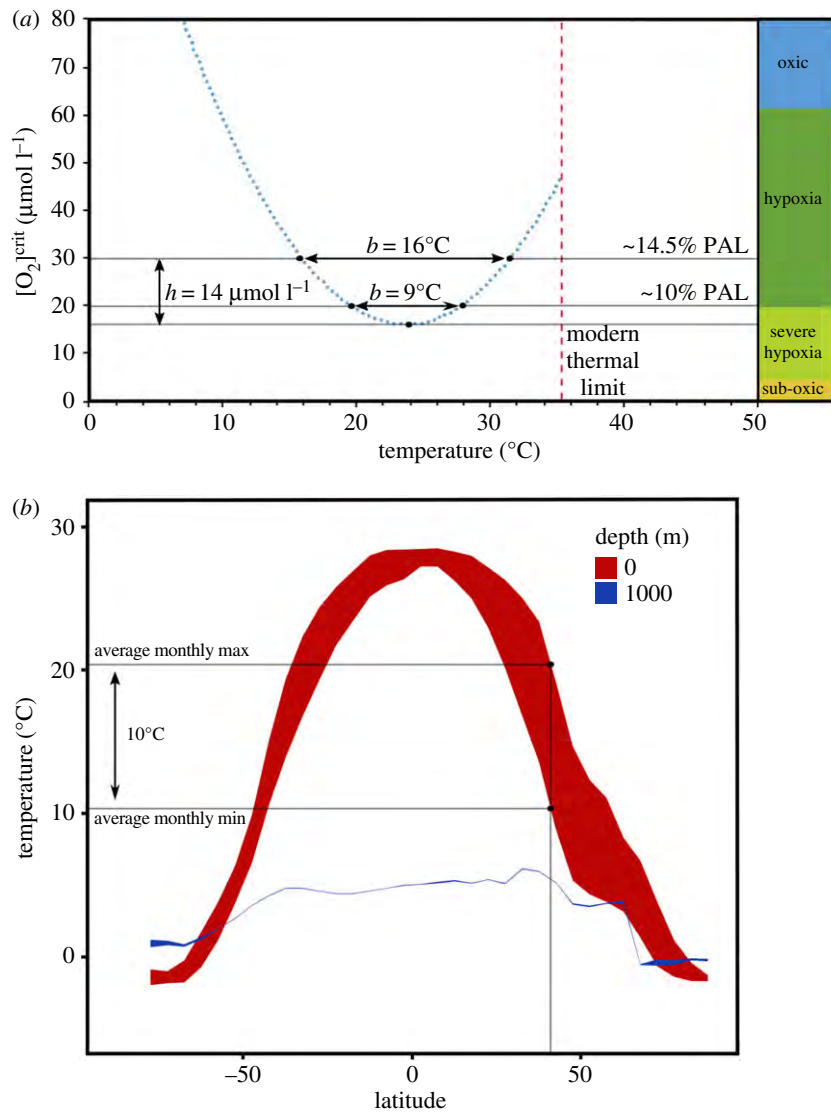
known about exact atmospheric  $\text{O}_2$  levels during this time [5,7,8]. If  $p\text{O}_2$  were closer to the lower-end estimate of 1% PAL, then presumably all megascopic metazoans would be driven deep into the respective vertices of their OCLTT parabola-space (figure 4a). This is if any could have survived at all; while metazoan macrofauna (0.3 to approximately 50 mm in size) can often have lower  $\text{O}_2$  requirements [63], theoretical, experimental, and oceanographic evidence suggest minimum  $p\text{O}_2$  levels of 1–4% PAL are needed for non-bilaterian megafauna in the absence of thermal variability [64,68,69]. Furthermore, marine animals require sustained metabolic rates to be a factor of approximately 2 to 5 greater than resting demand in order to sustain ecological activity [40]. Thus, the thermal range of low  $p\text{O}_2$  tolerance in the Ediacaran may have been considerably narrow. We note that although this discussion is couched in terms of traditional Ediacaran  $\text{O}_2$  estimates, the general principle of lower thermal range holds for any degree of low-oxygen Earth system.

With low ambient  $\text{O}_2$  narrowing the thermal range of taxa, seasonal temperature extremes in the shallow Ediacaran surface ocean probably had a significant impact on the aerobic respiration of metazoans. The effects of seasonality in the modern ocean can be used as an example. In the mid-latitudes, average annual sea surface temperature ranges greater than 10°C at 40° N. By contrast, seasonal temperature variation in the deep ocean is minimal (less than 1°C) across almost all latitudes at 1000 m depth (figure 4b). We propose this difference ultimately governed metabolically viable habitat for early animals in the Ediacaran.

### (b) A stenothermal origin for the Ediacara biota

The Cold Cradle hypothesis [21] originally posited that the Ediacara biota evolved in shallow, cold-water environments owing to increased  $\text{O}_2$  solubility. However, we have shown that  $\text{O}_2$  bioavailability in the global ocean maintains a positive linear relationship with temperature (electronic supplementary material, figure S7d). Furthermore, although cold-water has been theorized to increase aerobic scope by reducing standard metabolic rate in well-oxygenated conditions, low  $p\text{O}_2$  environments negate any benefit that this provides [39]. At a whole-organism scale in the Ediacaran, aerobic scope throughout the global ocean was likely to be very limited if estimates for a lower  $\text{O}_2$  Earth system are correct. Given the interaction of temperature on  $\text{O}_2$  supply and aerobic demand, shallow-water environments would not have had the ideal ecophysiological conditions needed to establish diverse ecosystems given seasonal temperature fluctuations.

We hypothesize it was the lack of thermal variability, and not necessarily cold conditions, that can best explain the deep-water origin of Ediacara biota. These environments would have been far below the thermocline (roughly 200 m in the modern ocean), where the global ocean is cold and nearly isothermal (figure 4b). In such conditions, the synergistic effects of temperature and low environmental  $p\text{O}_2$  on aerobic scope are muted and organisms can instead optimize their biochemical functions through compensatory adaptation to a significant degree [41,70]. Despite the lower bioavailability of  $\text{O}_2$  in cold water, this compensatory capability allows modern animals to inhabit sub-zero waters in Antarctic habitats of the Southern Ocean [42], and severely undersaturated hypoxic deep-water OMZs on continental



**Figure 4.** Impact of seasonal temperature variation on aerobic respiration in low  $pO_2$  conditions. (a) Polynomial regression run through the  $[O_2]^{crit}$  field of *D. lineata* ( $R^2 = 0.75$ ). The region above this blue dashed line represents the temperature and  $O_2$  conditions in which this species can maintain aerobic respiration (note that the regression vertex does not perfectly correspond with the lowermost measured  $[O_2]^{crit}$  value). While the exact equation of the polynomial probably differs between species, the bidirectional nature of  $O_2$  tolerance with respect to temperature is probably universal [46]. Because the area of this aerobic field ( $A$ ) is dependent on the severity of hypoxia  $h$ , and the thermal range of the environment  $b$ , lower ambient concentrations of dissolved  $O_2$  result in a narrower thermal range of aerobic respiration. Black horizontal lines represent  $O_2$  levels at 14.5% PAL, 10% PAL, and the vertex of the parabola. The red dashed line is approximately  $35^{\circ}\text{C}$ , the average upper critical (lethal) long-term thermal limit of many shallow-water ectotherms living in modern tropical environments [66]. Descriptive low- $O_2$  state boundaries are adapted from [67]. (b) Average seasonal temperature ranges across all latitudes of the global ocean at surface and 1000 m depths. Envelope height (shown here for  $40^{\circ}\text{N}$ ) represents the difference between maximum and minimum mean monthly temperatures for a given latitude. Data from NOAA WOA13 V2. (Online version in colour.)

slopes [55]. In an ecophysiological context, this means that while ambient  $pO_2$  ( $h$  in figure 4a) governs the thermal range of aerobic tolerance (width of  $b$  within the parabola, figure 4a), animals are able to shift their thermal optimum,  $T_{opt}$  (vertex of figure 4a), to cooler temperatures via long-term adaptation [41]. The global distribution of Ediacaran genera [71], combined with the fact that multiple taxa were capable of inhabiting both deep- and shallow-water environments in the later Ediacaran, suggests that thermal acclimation capacity was unlikely to be an issue [33,72,73].

It would seem that stenothermal habitats (areas which experience only a narrow range of temperatures) at depth would be a key environmental attribute needed for animals evolving in a low  $pO_2$  world. While we hypothesize temperature variability may be the most important factor, the fact that such deep-water environments are colder than the surface

ocean is not inconsequential: this would be particularly beneficial for viable aerobic habitat if the global mid-Ediacaran upper ocean was significantly warmer than the modern. Today, animals in the tropics are already near their upper thermal limits that at ecological timescales are commonly approximately  $35^{\circ}\text{C}$  [43,45,66]. In other words, though warmer waters increases  $O_2$  bioavailability approximately linearly, the increase in metabolic rates and thus  $O_2$  demand with temperature is exponential, and in such conditions this quickly leads to temperature-induced hypoxia [40,45]. In this light, if there was an environmental control on the stratigraphic appearance of Ediacaran organisms, the up-slope movement onto the shelf and eventually into littoral habitats observed between the Avalon–White Sea and White Sea–Nama assemblages [31–33] could be read as either increased ambient  $pO_2$  or global cooling. A further

expectation of this model would be that if middle Ediacaran (ca 570 Ma) shallow-water fossil occurrences are discovered, we expect them to be much lower in abundance and diversity (i.e. stressed communities) than temporally equivalent deep-water communities.

## 5. Conclusion

The Ediacara biota appear to have originated in deep-water slope and basinal settings in a global ocean that was still dominated by widespread low  $pO_2$  conditions [5,12,33]. In marine settings, total dissolved  $O_2$  concentration is a linchpin for metazoan life, but the critical threshold values at which point it imposes limits on physiological function can vary dramatically depending on multiple environmental factors such as primary productivity and organic carbon respiration, pH, salinity, hydrostatic pressure, and especially temperature. As we demonstrate here for the first time to our knowledge, in marine environments, the bioavailability of  $O_2$  (OSI) cannot simply be inferred from absolute environmental  $O_2$  concentrations alone, but rather it is the product of solubility, partial pressure, and diffusivity together. This has important implications for previous hypotheses that have considered the effect of water temperature on  $O_2$  solubility in Ediacaran oceans [21], as diffusivity is actually a stronger lever on  $O_2$  bioavailability, and works in the opposite direction. As a result, it is energetically more costly for animals to respire in cold, viscous water, despite its greater  $O_2$  solubility [39]. Furthermore, with new experimental physiology data, we show that temperature and  $O_2$  are synergistic and together govern the aerobic scope of marine

ectothermic animals. The effects of temperature on  $O_2$  supply and demand cause  $O_2$  tolerance to vary widely as temperature fluctuates. As a result, Ediacaran environments which experienced a wide range of temperatures, such as shallow-water littoral zones and microbial bioherms, would have physiologically challenged early animal communities already living in low ambient  $O_2$  conditions. We hypothesize that the apparent deep-marine origin of the Ediacara biota in cold, stenothermal environments may be the evolutionary solution to  $O_2$  and temperature co-limitation.

**Data accessibility.** Data available from the Dryad Digital Repository: <http://dx.doi.org/10.5061/dryad.bf43443> [74].

**Authors' contributions.** T.H.B. and E.A.S. designed the study. T.H.B. and L.E.E. contributed physiological data to the analysis. R.G.S. and T.H.B. performed the analysis of oceanographic data and T.H.B. analysed the results. T.H.B. wrote the manuscript with input from R.G.S., L.E.E., P.M.H. and E.A.S.

**Competing interest.** We declare no competing interests.

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## References

- Rooney AD, Strauss JV, Brandon AD, Macdonald FA. 2015 A Cryogenian chronology: two long-lasting synchronous Neoproterozoic glaciations. *Geology* **43**, 459–462. (doi:10.1130/G36511.1)
- Macdonald FA *et al.* 2010 Calibrating the Cryogenian. *Science* **327**, 1241–1243. (doi:10.1126/science.1183325)
- Droser ML, Tarhan LG, Gehling JG. 2017 The rise of animals in a changing environment: global ecological innovation in the late Ediacaran. *Annu. Rev. Earth Planet. Sci.* **45**, 593–617. (doi:10.1146/annurev-earth-063016-015645)
- Scott C, Lyons TW, Bekker A, Shen Y, Poulton SW, Chu X, Anbar AD. 2008 Tracing the stepwise oxygenation of the Proterozoic ocean. *Nature* **452**, 456–459. (doi:10.1038/nature06811)
- Sperling EA, Wolock CJ, Morgan AS, Gill BC, Kunzmann M, Halverson GP, Macdonald FA, Knoll AH, Johnston DT. 2015 Statistical analysis of iron geochemical data suggests limited late Proterozoic oxygenation. *Nature* **523**, 451–454. (doi:10.1038/nature14589)
- Hardisty DS *et al.* 2017 Perspectives on Proterozoic surface ocean redox from iodine contents in ancient and recent carbonate. *Earth Planet. Sci. Lett.* **463**, 159–170. (doi:10.1016/J.EPSL.2017.01.032)
- Holland HD. 2006 The oxygenation of the atmosphere and oceans. *Phil. Trans. R. Soc. B* **361**, 903–915. (doi:10.1098/rstb.2006.1838)
- Lyons TW, Reinhard CT, Planavsky NJ. 2014 The rise of oxygen in Earth's early ocean and atmosphere. *Nature* **506**, 307–315. (doi:10.1038/nature13068)
- Chen X *et al.* 2015 Rise to modern levels of ocean oxygenation coincided with the Cambrian radiation of animals. *Nat. Commun.* **6**, 7142. (doi:10.1038/ncomms8142)
- Sahoo SK, Planavsky NJ, Jiang G, Kendall B, Owens JD, Wang X, Shi X, Anbar AD, Lyons TW. 2016 Oceanic oxygenation events in the anoxic Ediacaran ocean. *Geobiology* **14**, 457–468. (doi:10.1111/gbi.12182)
- Miller AJ, Strauss JV, Halverson GP, Macdonald FA, Johnston DT, Sperling EA. 2017 Tracking the onset of Phanerozoic-style redox-sensitive trace metal enrichments: new results from basal Ediacaran post-glacial strata in NW Canada. *Chem. Geol.* **457**, 24–37. (doi:10.1016/j.chemgeo.2017.03.010)
- Tostevin R *et al.* 2016 Low-oxygen waters limited habitable space for early animals. *Nat. Commun.* **7**, 12818. (doi:10.1038/ncomms12818)
- Wallace MW, Hood A, Shuster A, Greig A, Planavsky NJ, Reed CP. 2017 Oxygenation history of the Neoproterozoic to early Phanerozoic and the rise of land plants. *Earth Planet. Sci. Lett.* **466**, 12–19. (doi:10.1016/j.epsl.2017.02.046)
- Krause AJ, Mills BJW, Zhang S, Planavsky NJ, Lenton TM, Poulton SW. 2018 Stepwise oxygenation of the Paleozoic atmosphere. *Nat. Commun.* **9**, 4081. (doi:10.1038/s41467-018-06383-y)
- Reinhard CT, Planavsky NJ, Olson SL, Lyons TW, Erwin DH. 2016 Earth's oxygen cycle and the evolution of animal life. *Proc. Natl Acad. Sci. USA* **113**, 8933–8938. (doi:10.1073/pnas.1521544113)
- Nursall JR. 1959 Oxygen as a prerequisite to the origin of the Metazoa. *Nature* **183**, 1170–1172. (doi:10.1038/1831170b0)
- Cloud Jr PE. 1968 Atmospheric and hydrospheric evolution on the primitive Earth. *Science* **160**, 729–736. (doi:10.2307/1724303)
- Knoll AH, Carroll SB. 1999 Early animal evolution: emerging views from comparative biology and geology. *Science* **284**, 2129–2137. (doi:10.1126/science.284.5423.2129)
- Canfield DE, Poulton SW, Narbonne GM. 2007 Late-Neoproterozoic deep-ocean oxygenation and the rise of animal life. *Science* **315**, 92–95. (doi:10.1126/science.1135013)



20. Butterfield NJ. 2018 Oxygen, animals and aquatic bioturbation: an updated account. *Geobiology* **16**, 3–16. (doi:10.1111/gbi.12267)
21. Fedonkin MA. 2003 The origin of the Metazoa in the light of the Proterozoic fossil record. *Paleontol. Res.* **7**, 9–41. (doi:10.2517/prpsj.7.9)
22. Noble SR, Condon DJ, Carney JN, Wilby PR, Pharaoh TC, Ford TD. 2015 U-Pb geochronology and global context of the Charnian Supergroup, UK: constraints on the age of key Ediacaran fossil assemblages. *Geol. Soc. Am. Bull.* **127**, 250–265. (doi:10.1130/B31013.1)
23. Liu AG, Kenchington CG, Mitchell EG. 2015 Remarkable insights into the paleoecology of the Avalonian Ediacaran macrobiota. *Gondwana Res.* **27**, 1355–1380. (doi:10.1016/j.gr.2014.11.002)
24. Pu JP, Bowring SA, Ramezani J, Myrow P, Raub TD, Landing E, Mills A, Hodgins E, Macdonald FA. 2016 Dodging snowballs: geochronology of the Gaskiers glaciation and the first appearance of the Ediacaran biota. *Geology* **44**, 955–958. (doi:10.1130/G38284.1)
25. Boag TH, Darroch SAF, Laflamme M. 2016 Ediacaran distributions in space and time: testing assemblage concepts of earliest macroscopic body fossils. *Paleobiology* **42**, 574–594. (doi:10.1017/pab.2016.20)
26. Xiao S, Laflamme M. 2009 On the eve of animal radiation: phylogeny, ecology and evolution of the Ediacara biota. *Trends Ecol. Evol.* **24**, 31–40. (doi:10.1016/j.tree.2008.07.015)
27. Hoyal CJF, Han J. 2018 Cambrian petalonamid Stromatoveris phylogenetically links Ediacaran biota to later animals. *Palaentology* **61**, 813–823. (doi:10.1111/pala.12393)
28. Sperling EA, Peterson KJ, Laflamme M. 2011 Rangeomorphs, Thectardis (Porifera?) and dissolved organic carbon in the Ediacaran oceans. *Geobiology* **9**, 24–33. (doi:10.1111/j.1472-4669.2010.00259.x)
29. Waggoner B. 1999 Biogeographic analyses of the Ediacara biota: a conflict with paleotectonic reconstructions. *Paleobiology* **25**, 440–458. (doi:10.1017/S0094837300020315)
30. Wood DA, Dalrymple RW, Narbonne GM, Gehling JG, Clapham ME. 2003 Paleoenvironmental analysis of the late Neoproterozoic Mistaken Point and Trepassy formations, southeastern Newfoundland. *Can. J. Earth Sci.* **40**, 1375–1391. (doi:10.1139/e03-048)
31. Macdonald FA, Strauss JV, Sperling EA, Halverson GP, Narbonne GM, Johnston DT, Kunzmann M, Schrag DP, Higgins JA. 2013 The stratigraphic relationship between the Shuram carbon isotope excursion, the oxygenation of Neoproterozoic oceans, and the first appearance of the Ediacara biota and bilaterian trace fossils in northwestern Canada. *Chem. Geol.* **362**, 250–272. (doi:10.1016/j.chemgeo.2013.05.032)
32. Grazhdankin D. 2014 Patterns of evolution of the Ediacaran soft-bodied biota. *J. Paleontol.* **88**, 269–283. (doi:10.1666/13-072)
33. Narbonne GM, Laflamme M, Trusler PW, Dalrymple RW, Greentree C. 2014 Deep-water Ediacaran fossils from northwestern Canada: taphonomy, ecology, and evolution. *J. Paleontol.* **88**, 207–223. (doi:10.1666/13-053)
34. Jablonski D, Sepkoski JJ, Bottjer DJ, Sheehan PM. 1983 Onshore-offshore patterns in the evolution of Phanerozoic shelf communities. *Science* **222**, 1123–1125. (doi:10.1126/science.222.4628.1123)
35. Dejours P. 1989 From comparative physiology of respiration to several problems of environmental adaptations and to evolution. *J. Physiol.* **410**, 1–19. (doi:10.1113/jphysiol.1989.sp017517)
36. Benson BB, Krause D. 1984 The concentration and isotopic fractionation of oxygen dissolved in freshwater and seawater in equilibrium with the atmosphere. *Limnol. Oceanogr.* **29**, 620–632. (doi:10.4319/lo.1984.29.3.0620)
37. Piiper J, Dejours P, Haab P, Rahn H. 1971 Concepts and basic quantities in gas exchange physiology. *Respir. Physiol.* **13**, 292–304. (doi:10.1016/0034-5687(71)90034-X)
38. Hofmann AF, Peltzer ET, Brewer PG. 2013 Kinetic bottlenecks to respiratory exchange rates in the deep-sea. Part 1: oxygen. *Biogeosciences* **10**, 5049–5060. (doi:10.5194/bg-10-5049-2013)
39. Verberk WCEP, Bilton DT, Calosi P, Spicer JJ. 2011 Oxygen supply in aquatic ectotherms: partial pressure and solubility together explain biodiversity and size patterns. *Ecology* **92**, 1565–1572. (doi:10.1890/10-2369.1)
40. Deutsch C, Ferrel A, Seibel B, Pörtner HO, Huey RB. 2015 Climate change tightens a metabolic constraint on marine habitats. *Science* **348**, 1132–1135. (doi:10.1126/science.aaa1605)
41. Somero GN, Lockwood BL, Tomanek L. 2017 *Biochemical adaptations: response to environmental challenges from life's origins to the Anthropocene*, 1st edn. Sunderland, MA: Sinauer Associates.
42. Peck LS, Webb KE, Bailey DM. 2004 Extreme sensitivity of biological function to temperature in Antarctic marine species. *Funct. Ecol.* **18**, 625–630. (doi:10.1111/j.0269-8463.2004.00903.x)
43. Sunday JM, Bates AE, Dulvy NK. 2012 Thermal tolerance and the global redistribution of animals. *Nat. Clim. Chang.* **2**, 686–690. (doi:10.1038/ndimate1539)
44. Sokolova IM, Frederich M, Bagwe R, Lannig G, Sukhotin AA. 2012 Energy homeostasis as an integrative tool for assessing limits of environmental stress tolerance in aquatic invertebrates. *Mar. Environ. Res.* **79**, 1–15. (doi:10.1016/J.MARENRES.2012.04.003)
45. Pörtner HO, Knust R. 2007 Climate change affects marine fishes through the oxygen limitation of thermal tolerance. *Science* **315**, 95–97. (doi:10.1126/science.1135471)
46. Pörtner H-O. 2010 Oxygen- and capacity-limitation of thermal tolerance: a matrix for integrating climate-related stressor effects in marine ecosystems. *J. Exp. Biol.* **213**, 881–893. (doi:10.1242/jeb.037523)
47. Melzner F, Bock C, Pörtner H-O. 2006 Critical temperatures in the cephalopod *Sepia officinalis* investigated using *in vivo* 31P NMR spectroscopy. *J. Exp. Biol.* **209**, 891–906. (doi:10.1242/jeb.02054)
48. Melzner F, Bock C, Pörtner HO. 2006 Temperature-dependent oxygen extraction from the ventilatory current and the costs of ventilation in the cephalopod *Sepia officinalis*. *J. Comp. Physiol. B* **176**, 607–621. (doi:10.1007/s00360-006-0084-9)
49. Vaquer-Sunyer R, Duarte CM. 2011 Temperature effects on oxygen thresholds for hypoxia in marine benthic organisms. *Glob. Chang. Biol.* **17**, 1788–1797. (doi:10.1111/j.1365-2486.2010.02343.x)
50. Zielinski S, Pörtner HO. 1996 Energy metabolism and ATP free-energy change of the intertidal worm *Sipunculus nudus* below a critical temperature. *J. Comp. Physiol. B* **166**, 492–500. (doi:10.1007/s003600050037)
51. Kristensen E. 1983 Ventilation and oxygen uptake by three species of *Nereis* (Annelida: Polychaeta). I. Effects of hypoxia. *Mar. Ecol. Prog. Ser.* **12**, 289–297. (doi:10.3354/meps012289)
52. Perry SF, Jonz MG, Gilmour KM. 2009 Oxygen sensing and the hypoxic ventilatory response. *Fish Physiol.* **27**, 193–253. (doi:10.1016/S1546-5098(08)00005-8)
53. Hofmann AF, Peltzer ET, Walz PM, Brewer PG. 2011 Hypoxia by degrees: establishing definitions for a changing ocean. *Deep Sea Res. Part I* **58**, 1212–1226. (doi:10.1016/j.dsr.2011.09.004)
54. Enns T, Scholander PF, Bradstreet ED. 1965 Effect of hydrostatic pressure on gases dissolved in water. *J. Phys. Chem.* **69**, 389–391. (doi:10.1021/j100886a005)
55. Levin L. 2003 Oxygen minimum zone benthos: adaptation and community response to hypoxia. *Oceanogr. Mar. Biol. Annu. Rev.* **41**, 1–45.
56. St-Denis CE, Fell CJD. 1971 Diffusivity of oxygen in water. *Can. J. Chem. Eng.* **49**, 885. (doi:10.1002/cjce.5450490632)
57. Spicer JJ, Gaston KJ. 1999 Amphipod gigantism dictated by oxygen availability? *Ecol. Lett.* **2**, 397–403. (doi:10.1046/j.1461-0248.1999.00105.x)
58. Somero GN. 2002 Thermal physiology and vertical zonation of intertidal animals: optima, limits, and costs of living. *Integr. Comp. Biol.* **42**, 780–789. (doi:10.1093/icb/42.4.780)
59. Liu AG, Matthews JJ, Menon LR, Mclroy D, Brasier MD. 2014 *Haoitia quadriformis* n. gen., n. sp., interpreted as a muscular cnidarian impression from the Late Ediacaran period (approx. 560 Ma). *Proc. R. Soc. B* **281**, 20141202. (doi:10.1098/rspb.2014.1202)
60. Liu AG, Mclroy D, Brasier MD. 2010 First evidence for locomotion in the Ediacara biota from the 565 Ma Mistaken Point Formation, Newfoundland. *Geology* **38**, 123–126. (doi:10.1130/G30368.1)
61. Gehling JG. 1988 A cnidarian of actinian-grade from the Ediacaran Pound Subgroup, South Australia. *Alcheringa* **12**, 299–314. (doi:10.1080/03115518808619129)
62. Sommer A, Klein B, Pörtner HO. 1997 Temperature induced anaerobiosis in two populations of the polychaete worm *Arenicola marina* (L.). *J. Comp. Physiol. B* **167**, 25–35. (doi:10.1007/s003600050044)

63. Sperling EA, Halverson GP, Knoll AH, Macdonald FA, Johnston DT. 2013 A basin redox transect at the dawn of animal life. *Earth Planet. Sci. Lett.* **371–372**, 143–155. (doi:10.1016/J.EPSL.2013.04.003)
64. Mills DB, Ward LM, Jones C, Sweeten B, Forth M, Treusch AH, Canfield DE. 2014 Oxygen requirements of the earliest animals. *Proc. Natl Acad. Sci. USA* **111**, 4168–4172. (doi:10.1073/pnas.1400547111)
65. Sperling EA, Frieder CA, Raman AV, Girguis PR, Levin LA, Knoll AH. 2013 Oxygen, ecology, and the Cambrian radiation of animals. *Proc. Natl Acad. Sci. USA* **110**, 13 446–13 451. (doi:10.1073/pnas.1312778110)
66. Nguyen KDT, Morley SA, Lai CH, Clark MS, Tan KS, Bates AE, Peck LS. 2011 Upper temperature limits of tropical marine ectotherms: global warming implications. *PLoS ONE* **6**, e29340. (doi:10.1371/journal.pone.0029340)
67. Sperling EA, Knoll AH, Girguis PR. 2015 The ecological physiology of Earth's second oxygen revolution. *Annu. Rev. Ecol. Syst.* **46**, 215–235. (doi:10.1146/annurev-ecolsys-110512-135808)
68. Runnegar B. 1991 Precambrian oxygen levels estimated from the biochemistry and physiology of early eukaryotes. *Palaeogeogr. Palaeoclimatol. Palaeoecol.* **97**, 97–111. (doi:10.1016/0031-0182(91)90186-U)
69. Levin LA, Huggett CL, Wishner KF. 1991 Control of deep-sea benthic community structure by oxygen and organic gradients in the eastern Pacific Ocean. *J. Mar. Res.* **49**, 763–800.
70. White CR, Alton LA, Frappell PB. 2012 Metabolic cold adaptation in fishes occurs at the level of whole animal, mitochondria and enzyme. *Proc. R. Soc. B* **279**, 1740–1747. (doi:10.1098/rspb.2011.2060)
71. Bowyer F, Wood RA, Poulton SW. 2017 Controls on the evolution of Ediacaran metazoan ecosystems: a redox perspective. *Geobiology* **15**, 516–551. (doi:10.1111/gbi.12232)
72. Gehling JG, Droser ML. 2013 How well do fossil assemblages of the Ediacara biota tell time? *Geology* **41**, 447–450. (doi:10.1130/G33881.1)
73. Grazhdankin DV, Balthasar U, Nagovitsin KE, Kochnev BB. 2008 Carbonate-hosted avalon-type fossils in arctic Siberia. *Geology* **36**, 803–806. (doi:10.1130/G24946A.1)
74. Boag TH, Stockey RG, Elder LE, Hull PM, Sperling EA. 2018 Data from: Oxygen, temperature, and the deep-marine stenothermal cradle of Ediacaran evolution. Dryad Digital Repository. (<http://dx.doi.org/10.5061/dryad.bf43443>)